# RESEARCH

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# Association between blood eosinophil count and Duchenne muscular dystrophy severity and prognosis: a retrospective cohort study



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# Abstract

Background Duchenne muscular dystrophy (DMD) is a rare hereditary muscular disease. The role of eosinophils in DMD has not been clarified. This study aims to evaluate the association between peripheral blood eosinophil count and severity and prognosis of DMD.

Methods A retrospective cohort study was performed for 145 DMD patients between January 2012 and December 2020. Clinical data of 150 healthy children were collected as a control group. Logistic regression and Cox regression analyses were used to explore the influences of eosinophil count on DMD severity and prognosis.

**Results** Eosinophil count in DMD group was lower than the control group (Z = 2.163, P = 0.031). It was negatively correlated with Vignos scale score, Spearman correlation coefficient was p = 0.245, P = 0.040 (at admission), p = 0.137, P = 0.032 (at follow-up); was a protective factor for high Vignos scale score at admission [odds ratio (OR) = 0.038, 95%Cl: 0.002–0.752, P = 0.032] and follow-up (OR = 0.033,95%Cl: 0.001–0.121, P = 0.039). The Cox regression analysis indicated that elevated eosinophil count was correlated with better therapeutic efficacy for DMD patients [hazard ratio (HR) = 2.218, 95%CI: 1.154-3.924, P = 0.016].

**Conclusion** Eosinophil count in peripheral blood was correlated with the severity of DMD. It could indicate the therapeutic efficacy and prognosis of DMD patients to a certain extent. Eosinophils may be a potentially valuable biomarker or therapeutic target for DMD.

Keywords Eosinophils, Duchenne muscular dystrophy, Inflammation, Prognosis, Therapeutic efficacy

Note: This article has been included in the preprint website at https://www. researchsquare.com/; https://doi.org/10.21203/rs.3.rs-1383155/v2. We have made some revise to this article, so the content is some different from that on the preprint website (the language of this article has been edited. Some tables and figures have been modified).

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## Background

Duchenne muscular dystrophy (DMD) is a severe X-linked disease characterized by progressive muscle weakness [1, 2]. Treatment of DMD is always a clinical challenge. The pathology of DMD involves gene abnormality, oxidative stress [3-5]. Inflammation, metabolic abnormalities, autophagy and regeneration defects are the major causes of it [6]. Inflammation plays a crucial role in the progression of DMD. Regulating inflammatory response and inducing immune tolerance to expression of dystrophin are key to treatment of DMD [7]. Nuclear factor-KB (NF-KB) signaling pathway promotes



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Eosinophils contribute to initiation and modulation inflammation.

It may be caused by allergy, infectious, inflammatory, neoplastic disorders. Eosinophils regulate immune homeostasis, inhibiting proinflammatory response of overreaction by secreting specific molecules [10]. Eosinophils in peripheral blood are associated with the prognosis of chronic obstructive pulmonary disease (COPD), infectious diseases, several cancer [11–13].

Study confirmed eosinophil infiltration in muscle tissue of DMD patients. It can promote repair muscle cells [14]. On the other hand, persistence of eosinophils within dystrophic muscle would sustain fibroblast proliferation, which promotes fibrosis tissue deposition, accelerating clinical decline of disease [2]. The role of eosinophils in DMD is still controversial and unclear. Few studies to explore whether it affects long-term outcome of DMD, to date. The present study aimed to assess the association between the eosinophil count in peripheral blood and the severity and prognosis of DMD.

## Methods

## Study design

Our cohort comprised children with DMD from Hunan Children's Hospital (Changsha, China) from January 1, 2012 to December 31, 2020. The inclusion criteria were as follows. 1) Boys aged within 2-5 years old (or slightly lower than 2 years old) with typical clinic-al symptoms. 2) Confirmation of DMD by genetic testing. 3) They had not been dealt at admission. The exclusion criteria were as follows: 1) Genetic testing exclude DMD. 2) Basic diseases, including infection, diarrhea, allergy, immune deficiency disease, or blood system disease. 3) Children with incomplete follow-up data. 4) Follow up was not completed. In total, 150 healthy children were enrolled as a control group. The inclusion criteria for the control group were as follows: 1) Healthy boys without obvious diseases during 3 months of follow-up. 2) Voluntary participation in vaccination or physical examination in our hospital. 3) Healthy volunteers participating in the valuedriven health plan for testing of blood and other biochemical indicators in our hospital and Beijing Children's Hospital. The exclusion criteria for the control group were as follows: 1) Combined with some diseases, such as infection, diarrhea, allergy. 2) Children with developmental or metabolic abnormalities. 3) Children with diseases that might influence the clinical data. 4) Study withdrawal due to various reasons. The present study was approved by the Ethics Committee of Hunan Children's Hospital (Approval No. KS2022-15).

## Data collection

Clinical data were collected for age and muscle strength score (Vignos scale [3]). All laboratory data, such as blood routine and myocardial enzyme spectrum were collected. While vitamin D3 (at admission, n = 55; at follow-up, n = 46, note: vitamin D examination of the control group and case group was matched by season) and C-reactive protein (CRP; at admission, n = 62; at follow-up, n = 41) were collected only from some children. All blood routine and biochemical tests were performed at 6:30–8:30 a.m.

## Treatment

- 1) Conservative treatment (n = 50): When children's parents knowing DMD was incurable, some parents declined prednisone treatment and selected conservative treatment, such as fructose sodium diphosphate and coenzyme Q10 supplement. A few children's parents did not decide whether receive conservative treatment during the treatment period, while they were followed up regularly (n = 6).
- Prednisone therapy (n = 95): Children aged within 5 years old received prednisone monotherapy (0.75 mg/kg/d QD), some children slightly late received this drug. Children were routinely supplemented calcium and vitamin D.

## Follow-up

Patients were followed up regularly after hospital discharge. They returned to the outpatient clinic of our hospital every 3–6 months to evaluate muscle strength and disease progression. The follow-up was terminated on April 30, 2021. The adverse reactions of prednisone and other drugs were monitored regularly. The therapeutic effects were evaluated at the last follow-up, the blood routine, myocardial enzymes were rechecked.

#### Statistical analysis

Normally distributed data were expressed as mean ± standard deviation. Data with skewed distribution were presented as median (P25,P75). Normally distributed data were analyzed by independent-samples t-test or paired t-test. The Mann–Whitney U test was utilized for analysis of abnormally distributed data. Pearson correlation analysis was used for assessment of the relationship between normally distributed data, Spearman correlation analysis was employed for abnormally distributed data. The logistic regression analysis was applied to evaluate the risk factors. The Kaplan–Meier analysis and Cox regression analysis were utilized to explore effects of various factors on the treatment efficacy. All data were processed using SPSS 24.0 software (IBM, Armonk, NY, USA). P < 0.05 was considered statistically significant.

## Results

## Patient clinical profiles

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Among 287 children were primarily enrolled, 80 cases did not complete blood routine and muscle enzyme tests. Forty cases lost follow-up because of not followed up on time or changed contact information. Twenty-two cases withdrew from the study because of pulmonary infection or diarrhea during treatment or follow-up. Thus, 145 children were involved in our cohort, and 150 children were included in the control group. The eosinophil level was 0 for eosinophil count of  $0-0.16 \times 10^9$ /l. It was 1 when eosinophil count  $\geq 0.16 \times 10^9$ /l. The results of genetic testing showed that 145 cases completed genetic testing, including 92 cases of gene deletion (80 cases of large fragments and 12 cases of small fragments), 22 cases of gene duplication and variation (4 cases of single-exon duplication and 16 cases of multi-exon duplication), 31 cases of point mutation (4missense variants, 16 nonsense variants, 6 frameshift variants, 5 splice variants). The mean follow-up time in the DMD group was 1.67 (0.50, 3.25) years. The detailed information of DMD and control group is shown in Table 1. The number of eosinophils in the DMD group was significantly lower than that in control group (Fig. 1).

One hundred forty-five children with DMD completed follow-up, in all. Compared with data collected at admission, the eosinophil count, erythrocyte level, hemoglobin level at follow-up were significantly different (P<0.05 or 0.01). The results are shown in Table 2.

## **Correlation analysis**

There were correlations between eosinophil count and vitamin D3level, neutropil-to-lymphocyte ratio (NLR) in the control group (Spearman correlation coefficient p = 0.299, P = 0.012, p = -0.261, P = 0.018). The correlation coefficient between eosinophil count and Vitamin D3 level in DMD group at admission was p = 0.563 (P = 0.001), at follow-up was p = 0.267(P = 0.207).

There were correlations between the eosinophil count and CRP level, NLR at admission (p = 0.271, P = 0.046, p = 0.232, P = 0.027), at follow-up (p = 0.189, P = 0.035, p = 0.227, P = 0.023) in DMD group.

Correlation coefficient between eosinophil count and Vignos scale score at admission was p = 0.245 (P = 0.040). Patients were divided into conservative treatment group (n = 50) and prednisone treatment group (n = 95). In prednisone treatment group, the correlation coefficient between eosinophil count and Vignos scale score at admission was p = 0.259 (P = 0.041), at follow-up was p = 0.092 (P = 0.023). In conservative treatment group, correlation coefficient between eosinophil count and

Table 1 General characteristics of the DMD and	control group
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	DMD group N = 145	Control group N = 150	Value	Р
Age (years)	3.83 (1.42,6.08)	2.92 (1.17,5.92)	-0.795	0.427
Eosinophil count(×10 <sup>9</sup> /l)	0.16 (0.09, 0.29)	0.21 (0.12, 0.32)	2.163	0.031
Ratio of eosinophils	0.02 (0.01,0.03)	0.03 (0.02,0.05)	3.454	0.001
White blood cell count(×10 <sup>9</sup> /l)	8.00 (6.39, 9.82)	7.55 (6.40, 9.41)	0.880	0.379
Neutrophil count(× 10 <sup>9</sup> /l)	3.43 (2.52, 4.36)	2.87 (2.30, 3.86)	2.723	0.006
Lymphocyte count(× 10 <sup>9</sup> /l)	3.34 (2.69, 4.03)	3.47 (2.87, 4.73)	-1.407	0.159
Monocyte count(×10 <sup>9</sup> /l)	0.35 (0.27, 0.49)	0.41 (0.32, 0.59)	-2.896	0.004
Red blood cell count(×10 <sup>12</sup> /l)	$4.65 \pm 0.37$	$4.79 \pm 0.39$	3.060	0.002
Hemoglobin(g/l)	122.83±10.29	128.51±9.89	4.651	0.000
Basophil count(× 10 <sup>9</sup> /l)	0.01 (0.003, 0.02)	0.01 (0.002, 0.03)	0.940	0.347
Platelet count(×10 <sup>9</sup> /l)	311.24±92.23	$308.32 \pm 65.48$	0.297	0.767
Vitamin D3(nmol/l)	47.91±11.48	82.60±23.12	6.439	0.000
Total protein(g/l)	66.18±4.13	68.66±4.41	4.820	0.000
Albumin(g/l)	42.52±3.71	46.74±2.39	11.042	0.000
Globulin(g/l)	23.67±3.87	$21.93 \pm 3.02$	4.193	0.000
Creatinine (umol/l)	18.41±7.79	$35.25 \pm 10.18$	13.375	0.000
NLR	1.04 (0.71,1.55)	0.87 (0.60,1.21)	2.535	0.011

NLR neutropil-to-lymphocyte ratio

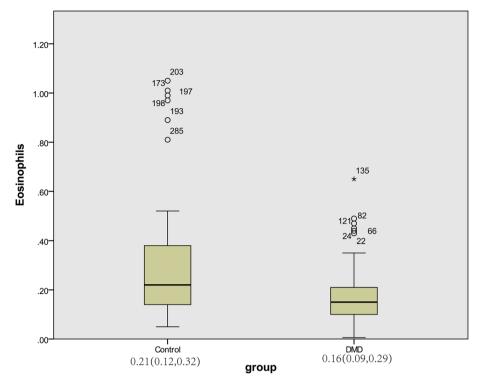




Fig. 1 Different eosinophil count between DMD and control group

 Table 2
 Clinical characteristics of DMD patients at admission and follow-up

	cases at admission ( <i>N</i> = 145)	cases at follow-up (N = 145)	Value	Р
Age (years)	3.83 (1.42,6.08)	5.25 (3.16,7.16)	10.449	0.000
Eosinophil count(× 10 <sup>9</sup> /l)	0.16 (0.09,0.29)	0.11 (0.07,0.19)	2.251	0.032
Ratio of eosinophils	0.02 (0.01,0.03)	0.02 (0.01,0.03)	1.105	0.269
White blood cell count(×10 <sup>9</sup> /l)	8.00 (6.39,9.82)	7.58 (6.16,8.87)	0.421?	0.674?
Basophil count(× 10 <sup>9</sup> /l)	0.01 (0.003,0.02)	0.01 (0.01,0.02)	0.766	0.444
Redblood cell count(×10 <sup>12</sup> /l)	4.65±0.37	4.85±0.42	4.577	0.000
Hemoglobin(g/l)	122.83±10.29	129.99±12.50	5.521	0.000
Neutrophil count(×10 <sup>9</sup> /l)	3.43 (2.52,4.36)	3.19 (2.71,4.41)	0.895	0.371
Lymphocyte count(× 10 <sup>9</sup> /l)	3.34 (2.69,4.03)	3.51 (2.51,4.27)	0.330	0.741
Monocyte count(×10 <sup>9</sup> /l)	0.35 (0.27,0.49)	0.40 (0.33,0.52)	-1.108	0.268
Platelet count(×10 <sup>9</sup> /l)	311.24±92.23	299.76±66.14	1.079	0.284
Vitamin D3(nmol/l)	$47.91 \pm 11.48$	45.22±9.23	1.155	0.307
Vignos scale	2 (1,3)	2 (1,3)	0.780	0.435
CK(U/L)	4177.00 (1698.70,9831.30)	7108.30 (1993.20,11,755.30)	2.074	0.038
Total protein(g/l)	66.18±4.13	66.24±4.29	0.425	0.672
Albumin(g/l)	42.52±3.71	42.62±3.35	0.049	0.961
Globulin(g/l)	23.67±3.87	23.62±2.97	0.398	0.692
CRP (mg/l)	0.75 (0.18,1.21)	0.50 (0.30,0.84)	1.521	0.128
Creatinine (umol/l)	18.41±7.79	23.41±9.17	2.933	0.006
NLR	1.02 (0.71,1.55)	0.95 (0.76,1.47)	1.333	0.182

CK Creatinekinase; CRP C-reactive protein; NLR neutropil-to-lymphocyte ratio

Vignos scale score at admission was p = 0.212 (P = 0.043), at follow-up was p = 0.233 (P = 0.004).

Correlation coefficient between lymphocyte count and Vignos scale score at admission was p=0.257, (P=0.004), at follow-up was p=0.195(P=0.032). The correlation coefficient between eosinophil count and lymphocyte count, monocyte count at admission was p=0.244 (P=0.007), p=0.182 (P=0.044); at follow-up was p=0.038 (P=0.7160), p=0.46 (P=0.656).

## **Effects of treatment**

In prednisone treatment group, eosinophil count was 0.15 (0.08, 0.25)×10<sup>9</sup>/l at time of treatment and 0.08 (0.05,0.17)×10<sup>9</sup>/l at follow-up. The eosinophil count was different statistically before and after treatment (Z=3.157, P=0.002).

The eosinophil count in the conservative treatment group was 0.19 (0.10, 0.31)×10<sup>9</sup>/l at admission, and 0.20 (0.11, 0.27)×10<sup>9</sup>/l at follow-up.

The eosinophil count was no difference before and after treatment (Z=0.270, P=0.787), statistically. The eosinophil count in conservative treatment group and prednisone treatment group was compared with that before treatment (Z=1.464, P=0.143), in which there was no statistically significant difference, while there was a significant difference after treatment (Z=4.559,P=0.000).

## Eosinophil affected muscle strength

Single-factor regression analysis of Vignos scale score in patients with DMD at admission revealed that age at admission, creatinine level, lymphocyte count, mononuclear count, eosinophil count was statistically significant. Age was a risk factor, while others were protective factors for muscle strength of DMD patients (Table 3).

In multivariate logistic regression analysis, we found that the eosinophil count [odds ratio (OR)=0.038, 95% confidence interval (CI): 0.002–0.752, P=0.032] was a protective factor for muscle strength score. The higher eosinophil count, the lower the score, and the higher muscle strength. Results of logistic regression analysis

of factors influencing muscle strength at admission are shown in Table 3.

The univariate analysis of Vignos scale score at followup revealed that age at follow-up, treatment, lymphocyte count, monocyte count, and Vignos scale score at admission were statistically significant. The eosinophil count at follow-up and at admission was also found statistically significant.

Further controlling of age and other factors performed by the multivariate regression analysis revealed the effects of treatment method (OR=0.167, 95%CI: 0.030-0.931, P=0.041) and Vignos scale score at admission (OR=13.582, 95% CI: 1.332-138.492, P=0.028). The effects of eosinophil count at follow-up on muscle strength score were statistically significant (OR=0.033, 95%CI: 0.001-0.821, P=0.039). Results of logistic regression analysis of factors influencing muscle strength during follow-up are presented in Table 4.

## Eosinophil affect therapeutic efficacy

The results of univariate Cox regression analysis revealed that age [hazard ratio (HR)=0.851, 95%CI: 0.771–0.939, P=0.001], treatment modality (HR=3.362, 95%CI:1.222–5.607, P=0.000), Vignos scale score (HR=0.637, 95%CI: 0.454–0.863, P=0.002), lymphocyte count (HR=1.056, 95%CI: 1.003–1.111, P=0.037), granulocyte count (HR=0.785, 95%CI: 0.457–0.954, P=0.048), albumin level (HR=1.131, 95%CI: 1.060–1.206, P=0.002) at admission; granulocyte count (HR=0.481, 95% CI: 0.254–0.912, P=0.025) and eosinophil count at follow-up (HR=1.895, 95%CI: 1.313–2.733, P=0.001) were statistically significant.

The multivariate regression analysis revealed that age (HR=0.929, 95% CI: 0.869–0.933, P=0.031) and Vignos scale score (HR=0.484, 95%CI: 0.267–0.880, P=0.017) at admission, treatment modality (HR=4.174, 95%CI: 2.401–7.495, P=0.003), eosinophil count at follow-up (HR=2.218, 95%CI: 1.154–3.924, P=0.016) were statistically significant. The results suggested that the higher

#### Table 3 Influencing factors for muscle strength score in DMD patients at admission

	Univariate analysis			Multivariat	Multivariate analysis		
	OR	95%CI	Р	OR	95%Cl	Р	
Age	1.453	1.180–1.789	0.000	2.619	1.369-5.003	0.004	
Lymphocyte count	0.646	0.434-0.960	0.031	0.531	0.237-1.190	0.124	
Eosinophil count	0.007	0.001-0.276	0.008	0.038	0.002-0.752	0.032	
Creatinine	0.865	0.754-0.953	0.039	0.950	0.889-1.106	0.135	
Monocyte count	0.046	0.003-0.849	0.038	0.191	0.022-1.653	0.133	

OR odds ratio; Cl Confidence interval

	Univariate analysis			Multivariate analysis		
	OR	95%CI	Р	OR	95%Cl	Р
Age	1.334	1.134–1.570	0.000	2.834	0.831-9.668	0.096
Treatment	0.404	0.178-0.918	0.030	0.167	0.030-0.931	0.041
Lymphocyte count	0.630	0.439-0.904	0.012	0.767	0.568-1.035	0.083
Monocyte count	0.015	0.001-0.405	0.013	0.169	0.016-1.739	0.135
Vignos scale score at admission	11.543	5.607-26.297	0.000	13.582	1.332-138.492	0.028
Eosinophil count at follow-up	0.012	0.001-0.645	0.029	0.033	0.001-0.821	0.039
Eosinophil count at admission	0.016	0.002-0.586	0.024	0.002	0.001-20.452	0.167

## Table 4 Influencing factors for muscle strength score in DMD patients during follow-up

OR odds ratio; CI Confidence interval

eosinophil count, the better therapeutic efficacy. These results are shown in Table 5.

Kaplan–Meier analysis of eosinophil count at follow-up showed that there were significant differences in different eosinophil count on survival time of children with DMD. The median survival time of 0 level was 1.083 years, standard deviation was 0.273, while that of 1 level was 2.524 years, standard deviation was 0.519 (Fig. 2).

The higher eosinophil count, the longer survival time of children with DMD. There were also significant differences in effects of different treatment modalities on survival time of children with DMD during follow-up. The median survival time in the conservative treatment group was 1.160 years, standard deviation was 0.184; the median survival time in prednisone treatment group was 4.580 years, standard deviation was 0.405 (Fig. 3). The therapeutic efficacy of prednisone treatment was significantly higher than conservative treatment. (To evaluate the therapeutic efficacy more conveniently, we considered that the treatment was ineffective if the Vignos scale

Table 5	Influencing fa	tors for therapeutic	efficacy of DMD	patients
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score increased. If survival time was long, the therapeutic efficacy would be better).

In the Cox regression analysis, there were significant differences in the therapeutic efficacy of different eosinophil count on children with DMD (Fig. 4). The higher the eosinophil count, the better the therapeutic efficacy.

## Discussion

We demonstrated that the eosinophil count was related to muscle strength and long-term treatment effect in DMD patients. Eosinophilcount was negatively correlated with the intensity of inflammatory response in vivo. These may stem from the immunoregulatory and muscle repair of eosinophils.

Growing evidence proved that inflammation influences DMD. Continuous inflammatory response aggravates severity of DMD [3, 9]. Chronic inflammation is a pathological feature of DMD. DMD patients are accompanied with the elevated levels of inflammatory factors, such asinterleukin-6 (IL-6), interferon- $\gamma$  (IFN- $\gamma$ ), tumor

	Univariate Cox regression			Multivariate Cox regression		
	HR	95%Cl	Р	HR	95%Cl	Р
Age atadmission	0.851	0.771-0.939	0.001	0.929	0.869-0.993	0.031
Treatment modality	3.362	1.222-5.607	0.000	4.174	2.401-7.495	0.003
Vignos scale score at admission	0.637	0.454-0.863	0.002	0.484	0.267-0.880	0.017
Lymphocyte count at admission	1.056	1.003-1.111	0.037	1.232	0.949-1.613	0.117
Granulocyte count at admission	0.785	0.457-0.954	0.048	0.604	0.278-1.295	0.174
Granulocyte count at follow-up	0.481	0.254-0.912	0.025	0.611	0.285-1.439	0.198
Albumin at admission	1.131	1.060-1.206	0.002	1.250	0.941-1.643	0.109
Eosinophil count at follow-up	1.895	1.313-2.733	0.001	2.218	1.154–3.924	0.016
Eosinophil count at admission	1.281	0.743-1.572	0.684	1.147	0.904-1.463	0.164

HR Hazard ratio; CI Confidence interval

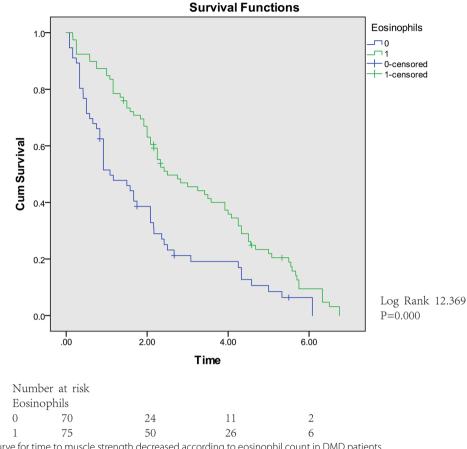


Fig. 2 Survival curve for time to muscle strength decreased according to eosinophil count in DMD patients

necrosis factor- $\alpha$  (TNF- $\alpha$ ), etc. The effectiveness of prednisone treatment is also a strong evidence of an inflammatory response.

There is a chronic inflammatory response in children with DMD. Eosinophils are engaged in immune inflammatory response. We confirmed that there was a certain correlation between eosinophil count and muscle strength score. It confirmed that eosinophil count had a definite impact on the muscle strength. We found that eosinophil count in children with DMD was negatively correlated with CRP level and neutropil-to-lymphocyte ratio (NLR). NLR is a simple inflammatory marker and is related to the prognosis of many diseases such as infection, tumor and immunity [15]. Therefore, eosinophil count was negatively correlated with systemic inflammatory response. Eosinophil can downregulate the inflammatory response. In our study, eosinophil count in the control group was higher than DMD group. This showed that the control group has higher ability to downregulate inflammatory response.

It is widely accepted that eosinophils are inflammatory cell, which is related to allergy and parasitic infection. The increase of eosinophil count represents an inflammatory response. Why is it negatively correlated with CRP level ? A COPD study confirmed that the eosinophil count in peripheral blood was negatively correlated with CRP level. That could be related to the fact that eosinophils enter the lung tissue when the inflammatory reaction is obvious, leading to eosinophils decreased [16]. We speculate that there is a similar reason, in which eosinophils enter muscle tissue, thus, eosinophil count in peripheral blood in DMD group was slightly lower than the control group. Eosinophils can maintain Th1/ Th2 (T helper1/T helper2) immune balance and inhibit excessive inflammatory response in vivo [17]. We believe that eosinophils inhibit the excessive inflammatory response in children with DMD. However, this needs more research to confirm it.

Our study showed that a high eosinophil count means better muscle strength and therapeutic efficacy (Fig. 2). However, previous study demonstrated that eosinophil count in peripheral blood was not related to eosinophil count in DMD muscle [18]. The relationship between its change and disease severity cannot be compared with the

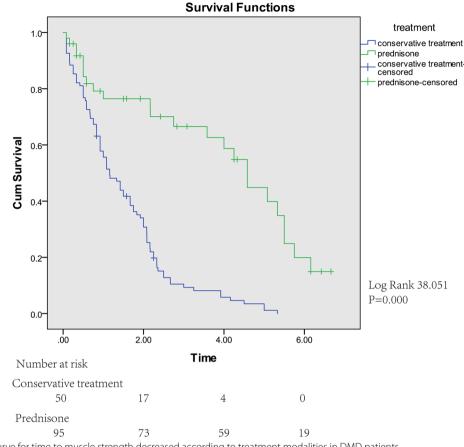


Fig. 3 Survival curve for time to muscle strength decreased according to treatment modalities in DMD patients

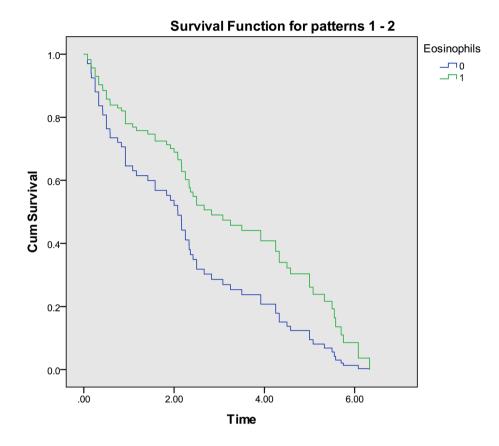
relationship between number of eosinophils in muscle and disease severity.

Our study revealed that the eosinophil count was negatively correlated with Vignos scale score before and after prednisone treatment, although it was very weak. This correlation persisted in the conservative treatment group. We found that eosinophils have a certain protective effect on muscle strength of DMD patients. However, the underlying mechanism should be further studied.

We concluded that eosinophils were protective factors for high motor function score whether at admission or at follow-up. After treatment, we found that the lower eosinophil count in prednisone treatment group, the worse therapeutic efficacy. Our Kaplan–Meier analysis and multivariate Cox regression analysis also supported this conclusion. The effects of eosinophil count on therapeutic efficacy of DMD might originate from its protective effects on muscle strength. Animal experiments demonstrated that eosinophils do not mediate the acute muscle injury [19], but has not been confirmed in human experiments. The present study showed that a high eosinophil count contributes to the improvement of long-term therapeutic efficacy of DMD. This may indirectly confirm that eosinophils play a certain role in repairing muscle damage in DMD patients [10]. However, this finding needs to be further verified.

Vitamin D regulates calcium and phosphorus metabolism as well as the inflammatory response. Its immunomodulatory ability has been used to treat multiple sclerosis and systemic lupus erythematous (SLE) [20, 21]. Vitamin D3 regulates the inflammatory response through the vitamin D receptor, enhance T regulatory (Treg) cell function, induce immune tolerance.

Our study confirmed that level of vitamin D3 in the DMD group was significantly lower than control group. Thus, the ability of DMD group to regulate the inflammatory response was relatively low. Our study revealed that the number of eosinophils was positively correlated with vitamin D3 level. Literature confirmed eosinophils inhibit excessive inflammatory response in vivo [17]. We speculate that eosinophils have similar ability to down-regulate inflammatory responses as vitamin D3. The stronger correlation between eosinophil count and vitamin D3 level in DMD group was due to lower vitamin



HR=2.218 95%CI=1.154-3.924 P=0.016

Fig. 4 Cox regression for independent factors on muscle strength decreased in DMD children after adjustment. Abbreviations: HR: hazard ratio CI: confidence interval

D3 level and lower eosinophil count in DMD group than control group.

Eosinophils are regulated by lymphocytes and their cytokines. T lymphocytes are involved in inflammatory response of DMD. Previous studies demonstrated that number of T cells decreased after prednisone treatment in DMD patients, which was accompanied by reduced muscle necrosis and fibrosis. Our study indicated that blood lymphocytes were protective factor of muscle strength scores at admission. However, multivariate Cox regression analysis showed no statistical significance. This could be due to the small sample size or other factors. Previous study reported that the number of CD8/ CD26T cells in peripheral blood of DMD patients was positively correlated with quantitative muscle score (i.e., the higher number, the greater muscle strength), which is consistent with our results [22]. Another research demonstrated that CD49d can be used as a marker and potential therapeutic target for DMD [23].

Eosinophils are activated by monocytes, which participating inflammatory response of DMD. Monocyte macrophages appear in the early-stage of muscle injury, in which pro-inflammatory macrophages mainly mediate the inflammatory response, and pro-regenerative macrophages inhibit this response to repair muscle cells [24]. Animal experiments have shown that inflammatory monocytes improve the prognosis of patients with DMD, and may play a role through chemokine receptor-2 (CCR2) [25]. Our study confirmed that the eosinophil count was correlated with monocyte changes. This may suggest that the role of eosinophils is similar.

Influence of age on Vignos scale score is obvious. For DMD, the deletion of dystrophin protein is caused genetically. With the increase of age, muscle damage, degradation, children with DMD may lose motor function at about 12 years old if do not timely receive treatment. With increase in age, the Vignos scale score gradually increases, which are consistent with our results. In our study, Cox regression analysis revealed that therapeutic efficacy was reduced with the increase of age. With increase in age, the interaction between chronic activation of innate immunity and degradation and regeneration of asynchrony and proximity produced an uncoordinated repair response. This repair response could promote the disease progression.

At present, effective treatment for DMD is prednisone, while its adverse reactions are noteworthy. To our knowledge, difecolate is more effective than prednisone (we did not use this drug in our study because few patients preferred to receive it) [26]. Although the treatment mechanism of prednisone for DMD has not been fully clarified [27, 28], our study confirmed that prednisone could control the inflammatory response. Our logistic regression suggested that prednisone had a protective effect on muscle strength. Muscle strength could be improved with prednisone treatment. Our Cox regression analysis further confirmed that prednisone was effective, which was consistent with previously reported findings [29].

We found eosinophil count can not only reflect inflammatory response of DMD patients, but also represent muscle strength or prognosis. Because some children cannot complete muscle strength test or 6-min walking test well. It is important to seek a simple and practical biomarker to indicate the muscle strength or degree of muscle injury in DMD patients. Eosinophil count can represent muscle strength to some extent. We confirmed eosinophil count affects therapeutic efficacy of children with DMD, so it can be used as a therapeutic target.

There were some deficiencies in the present study. First, the sample size was small, and the 6-min walk test was not completed (some children did not cooperate). Second, follow-up time of DMD children was not long enough. Third, the association of eosinophil count and levels of cytokines in children with DMD was not assessed. Fourth, this study did not investigate specific mechanisms. We need animal experiments to prove it.

## Conclusions

In conclusion, eosinophil count in peripheral blood of children with DMD could reflect muscle strength and inflammatory response, could be associated with therapeutic efficacy, could represent prognosis to a certain extent. Eosinophils may be a potentially valuable biomarker or therapeutic target for DMD.

#### Abbreviations

ORCID	Open Researcher and Contributor Identity Document
DMD	Duchenne muscular dystrophy
OR	Odds ratio
CI	Confidence interval
NF-ĸB	Nuclear factor-ĸB
COPD	Chronic obstructive pulmonary disease
CRP	C-reactive protein
QD	Quaque die
CK	Creatine kinase
NLR	Neutropil-to-lymphocyte ratio
IL-6	Interleukin-6

IFN-γ	Interferon-γ
TNF-α	Tumor necrosis factor-α
Th1	T helper1
Th2	T helper 2
SLE	Systemic lupus erythematous
Treg	T regulatory
CD8	Cluster of differentiation 8
CD26	Cluster of differentiation 26
QMT	Quantitative muscle score
CD49d	Cluster of differentiation 49d
CCR2	Chemokine receptor-2

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#### Authors' contributions

ZJ designed the study, extracted and analyzed data, drafted the manuscript, and prepared figures. HML revised the manuscript and participated in data collection. LWW directed the design of the research. WJH drew tables and reviewed data. LMY and BC participated in data collection and literature review. ZSN, JWT, XR, CM, and FG participated in drafting the manuscript. SLL performed the statistical analysis. All the authors read and approved the submitted version of the manuscript.

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#### Availability of data and materials

The data that support the findings of this study are available from the corresponding author upon reasonable request.

## Declarations

#### Ethics approval and consent to participate

This study was approved by the Ethics Committee of Hunan Children's Hospital (Approval No. KS2022-15). The informed consent was obtained from children's parents. We confirmed that all methods were carriedout in accordance with the ethical standards as laid down in the 2013 Declaration of Helsinki and its later amendments.

#### **Consent for publication**

Not applicable.

#### **Competing interests**

The authors declare that there is no competing interest.

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## References

- Gartz M, Lin CW, Sussman MA, Lawlor MW, Strande JL. Duchenne muscular dystrophy (DMD) cardiomyocyte-secreted exosomes promote the pathogenesis of DMD-associated cardiomyopathy. Dis Model Mech. 2020;13(11):dmm045559.
- Kastenschmidt JM, Coulis G, Farahat PK, Pham P, Rios R, Cristal TT, et al. A stromal progenitor and ILC2 niche promotes muscle eosinophilia and fibrosis associated gene expression. Cell Rep. 2021;35(2): 108997.
- Cruz-Guzmán Odel R, Rodríguez-Cruz M, Escobar Cedillo RE. Systemic Inflammation in Duchenne Muscular Dystrophy: Association with Muscle Function and Nutritional Status. Biomed Res Int. 2015;2015: 891972.
- Lecompte S, Abou-Samra M, Boursereau R, Noel L, Brichard SM. Skeletal muscle secretome in Duchenne muscular dystrophy: a pivotal antiinflammatory role of adiponectin. Cell Mol Life Sci. 2017;74(13):2487–501.
- Farini A, Sitzia C, Cassani B, Cassinelli L, Rigoni R, Colleoni F, et al. Therapeutic potential of immunoproteasome inhibition in Duchenne muscular dystrophy. MolTher. 2016;24(11):1898–912.

- Bello L, Pegoraro E. The, "Usual Suspects": Genes for inflammation, fibrosis, regeneration, and muscle strength modify Duchenne muscular dystrophy. J Clin Med. 2019;8(5):649.
- Villalta SA, Rosenberg AS, Bluestone JA. The immune system in Duchenne muscular dystrophy: Friend or foe. Rare Dis. 2015;3(1): e1010966.
- Rosenberg AS, Puig M, Nagaraju K, Hoffman EP, Villalta SA, Rao V, et al. Immune-mediated pathology in Duchenne muscular dystrophy. Sci Transl Med.2015;7(299):299rv294.
- Donovan JM, Zimmer M, Offman E, Grant T, & JirousekM. A novelNF-kappaB inhibitor, Edasalonexent (CAT-1004), in development as a diseasemodifying treatment for patients with Duchenne muscular dystrophy: Phase 1 safety, pharmacokinetics, and pharmacod-ynamicsin adult subjects. J Clin Pharmacol, 2017;57(5):627–639.
- Tidball JG, Welc SS, Wehling-Henricks M. Immunobiology of inherited muscular dystrophies. Compr Physiol. 2018;8(4):1313–56.
- 11. Moreira A, Leisgang W, Schuler G, Heinzerling L. Eosinophilic count as a biomarker for prognosis of melanoma patients and its importance in the response to immunotherapy. Immunotherapy. 2017;9(2):115–21.
- Fuschillo S, Molino A, Stellato C, Motta A, Maniscalco M. Blood eosinophils as biomarkers of therapeutic response to chronic obstructive pulmonary disease: still work in progress. Eur J Intern Med. 2019;68:1–5.
- Okauchi S, Shiozawa T, Miyazaki K, Nishino K, Sasatani Y, Ohara G, et al. Association between peripheral eosinophils and clinical outcomes in patients with non-small cell lung cancer treated with immune checkpoint inhibitors. Pol Arch Intern Med. 2021;131(2):152–60.
- Heredia JE, Mukundan L, Chen FM, Mueller AA, Deo RC, Locksley RM, et al. Type 2 innate signals stimulate fibro/adipogenic progenitors to facilitate muscle regeneration. Cell. 2013;153(3):376–88.
- Bartlett EK, Flynn JR, Panageas KS, Ferraro RA, Sta Cruz JM, Postow MA, et al. High neutrophil-to-lymphocyte ratio (NLR) is associated with treatment failure and death in patients who have melanoma treated with PD-1 inhibitor monotherapy. Cancer. 2020;126(1):76–85.
- Lv MY, Qiang LX, Li ZH, Jin SD. The lower the eosinophils, the stronger the inflammatory response? The relationship of different levels of eosinophils with the degree of inflammation in acute exacerbation chronic obstructive pulmonary disease (AECOPD). J Thorac Dis. 2021;13(1):232–43.
- Ondari E, Calvino-Sanles E, First NJ, Gestal MC. Eosinphils and bacteria, the beginning of a story. Int J Mol Sci. 2021;22(15):8004.
- Schröder T, Fuchss J, Schneider I, Stoltenburg-Didinger G, Hanisch F. Eosinophils in hereditary and inflammatory myopathies. Acta Myol. 2013;32(3):148–53.
- Sek AC, Moore IN, Smelkinson MG, Pak K, Minai M, Smith R, et al. Eosinophils do not drive a acute muscle pathology in the mdx mouse model of Duchenne muscular dystrophy. J Immunol. 2019;203(2):476–84.
- Lu M, Taylor BV, Körner H. Genomic effects of the vitamin D receptor: potentially the link between vitamin D, immune cells, and multiple sclerosis. Front Immunol. 2018;9:477.
- Mailhot G, White JH. Vitamin D and immunity in infants and children. Nutrients. 2020;12(5):1233.
- Soslow JH, Markham LW, Burnette WB, Galindo CL, Feoktistov I, Raucci FJ, et al. Increased number of circulating CD8/CD26T cellsinthe blood of Duchenne muscular dystrophy patients is associated with augmented binding of ddenosine deaminase and higher Muscular strength scores. Front Pharmacol. 2017;8:914.
- Pinto-Mariz F, Rodrigues Carvalho L, Prufer De Queiroz Campos Araujo A, De Mello W, Gonçalves Ribeiro M, Cunha Mdo C, et al. CD49d is a disease progression biomarker and a potential target for immunotherapy in Duchenne muscular dystrophy. Skeletal Muscle. 2015;5:45.
- Kharraz Y, Guerra J, Mann CJ, Serrano AL, Muñoz-Cánoves P. Macrophage plasticity and the role of inflammation in skeletal muscle repair. Mediators Inflamm. 2013;2013: 491497.
- Mojumdar K, Liang F, Giordano C, Lemaire C, Danialou G, Okazaki T, et al. Inflammatory monocytes promote progression of Duchenne muscular dystrophy and can be therapeutically targeted via CC-R2. EMBO Mol Med. 2014;6(11):1476–92.
- McDonaldCM, Sajeev G, Yao Z, McDonnell E, Elfring G, Souza M, et al. Deflazacort vs prednisone treatment for Duchenne muscular dystrophy: A meta-analysis of disease progression rates in recent multicenter clinical trials. Muscle Nerve. 2020;61(1):26–35.
- Flanigan KM, Campbell K, Viollet L, Wang W, Gomez AM, Walker CM, et al. Anti-Dystrophin T cell responses in Duchenne muscular dystrophy:

prevalence and a glucocorticoid treatment effect. Hum Gene Ther. 2013;24(9):797–806.

- Kameyama T, Ohuchi K, Funato M, Ando S, Inagaki S, Sato A. et al. Efficacy of prednisolone in generated myotubes derived from fibroblasts of Duchenne muscular dystrophy patients. Front Pharmacol. 2018; 9:1402.
- Herbelet S, Rodenbach A, Paepe B, DeBleecker JL. Anti-Inflammatory and general glucocorticoid physiology in skeletal muscles affected by Duchenne muscular dystrophy: exploration of steroid sparing agents. Int J Mol Sci. 2020;21(13):4596.

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